

The Journal of Anatomical Sciences Email: journalofanatomicalsciences@gmail.com J. Anat Sci 15(1) Effects of acute oral administration of aqueous extract of *Syzygium guineense* root on the kidney of albino Wistar rats

*Isa ZA, Garba SH, Zirahei JV, Attah MOO

Department of Human Anatomy, Faculty of Basic Medical Sciences, University of Maiduguri, P.M.B. 1069, Maiduguri, Borno State, Nigeria.

Corresponding author:

E-mail: zakariyaalhajii@gmail.com, Mobile Phone: +234 806 953 5737

Abstract

The first step in evaluating toxicity of a substance is to test for acute toxicity of that substance. The focus of acute oral toxicity is on possible short-term adverse effects from one or more oral exposures given within a 24-hour period. The purpose of this research was to investigate the effects of acute oral administration of *Syzygium guineense's* root extract on the kidney of albino Wistar rats. *S. guineense* belongs to the family *Myrtaceae*. It is used against cancer and stomach ache, and as antihelmintic and purgative. A total of fifteen (15) young healthy male Wistar rats (8-10 weeks old) weighing between 140 g and 165 g were used for this study. The study was designed and conducted in two (2) phases: phase I and phase II using established protocols. In phase I, twelve (12) rats were divided into four (I-IV) groups of three rats each. Group I was designated as the control group, while groups II, III and IV were administered with 10 mgkg⁻¹, 100 mgkg⁻¹ and 1000 mgkg⁻¹ respectively. In phase II, three rats were used and shared into three (I-III) groups of one rat each. Groups I, II and III were administered with 1600 mgkg⁻¹, 2900 mgkg⁻¹ and 5000 mgkg⁻¹ respectively. Data were analyzed using GraphPad Prism version 9.2.0. Results from this research were presented as mean±SEM. Relative kidney weights showed significant decrease. Plasma sodium and urea concentrations showed significant increase. Kidney sections from the treated groups showed no remarkable changes. It is recommended that the root of *S. guineense* should be used with caution.

Keywords: acute oral toxicity, Syzygium guineense, renal function, histology, albino Wistar rat

INTRODUCTION

Acute oral toxicity is crucial in assessing the safety of chemicals and pharmaceuticals, focusing on potential short-term negative effects from single oral exposure, a prevalent mode of exposure in real-world situations¹. Studies on acute toxicity offer insights on the possibility of acute toxicity in humans, the target organs of toxicity, the course of drug-induced clinical findings over time, the right dosage for multiple-dose toxicity studies, and the toxicity differences between species².

Syzygium guineense belongs to the family *Myrtaceae*. It is a tree that grows 10 to 15 meters high and has a thick, twisted bole, relatively low branching, a moderately dense crown, and drooping branches^{3. 4}. The whole plant has anticancer activity^{5,6}. *S. guineense* root infusion is used as an anthelmintic and purgative, and for the treatment of stomachache⁶. The leaf of *S. guineense* is used as anti-inflammatory, analgesic, antibacterial, antimalarial, antidiarrheal, antidiabetic (*in vitro*), antioxidant and

antihypertensive^{7, 8, 9}. The stem bark of *S. guineense* has antispasmodic activities and is used in the management of tuberculosis^{6, 10}.

Kidneys are two bean-shaped, smooth, reddish-brown organs, located retroperitoneally and are attached ventrally to the posterior abdominal wall¹¹. The kidneys are responsible for regulation of fluid, electrolyte and acid-base balance: detoxification: elimination of metabolic waste products and excretion of nitrogenous wastes such as creatinine and urea from the body. Estimates of the plasma or serum levels of certain compounds and electrolytes are typically used as biomarkers for kidney function. In addition, the kidneys serve endocrine functions^{12, 13, 14}. The functional unit of the kidney is called the nephron, which consists of renal corpuscle and uriniferous tubules¹⁵. The present research is aimed at investigating the acute oral effects of aqueous root extract of S. guineense on the kidney of albino Wistar rats.

MATERIALS AND METHODS

Collection and identification of S. guineense's roots

Fresh roots of S. guineense were obtained from a herb seller at Maiduguri Monday market in Borno State, Nigeria. The plant was identified and authenticated at the Department of Biological Sciences, University of Maiduguri, Nigeria, where a Voucher number UMDB001 was deposited at the herbarium. The plant material was dried under shade, pulverized and subjected to exhaustive soxhlet aqueous extraction. Continuous hot extraction is a procedure that uses a Soxhlet extractor, a glass-based device. The powdered root of S. guineense was placed in a porous bag, filled with water, and heated. The water evaporated, flowed through the condenser, and then into the extraction chamber, where it contacted the root of S. guineense to be extracted. The water level in the extraction chamber reached the top of the siphon, and the process repeated until all the drug is extracted.

Animal care

The rats were procured from the animal house of the Department of Biochemistry, Faculty of Science, University of Maiduguri, Nigeria. The research was also conducted in the same place. The rats had access to feed (Vital Feed Growers, Grand Cereals Ltd., Jos-Nigeria) and water *ad libitum*. Standard guidelines as required by the ethical committee of the University of Maiduguri for the care and use of laboratory animals was strictly followed throughout the research period.

Experimental design

With little modifications, the acute oral toxicity study was conducted according to Lorke's method¹⁶. A total of fifteen (15) young healthy male Wistar rats (8-10 weeks old) weighing between 140-165 g were used for this study. The study was designed and conducted in two (2) phases: phase I and phase II. In phase I, twelve (12) rats were divided into four (I-IV) groups of three rats each. Group I was designated as the control group, while groups II, III and IV were administered with 10 mgkg⁻¹, 100 mgkg⁻¹ and 1000 mgkg⁻¹ respectively. In phase II, three rats were used and shared into three (I-III) groups of one rat each. Groups I, II and III were administered with 1600 mgkg⁻¹, 2900 mgkg⁻¹ and 5000 mgkg⁻¹ respectively.

All experimental animals were dosed once. Phase I rats were observed for fourteen days. Starting from the first hour after treatment, they were observed on hourly basis for the first 24 hours, and then at approximately the same time on daily basis thereafter. Phase II rats were observed for a duration of 24 hours only after treatment, during that period, they were observed on hourly bases. At the end of the

experiment, the rats were euthanized by intraperitoneal injection of ketamine chloride 75 mg/kg (Hospira Inc. Lake Forest, USA). Blood samples were collected in plain bottles by cardiac puncture and centrifuged at 3500 rpm (using 896 MSE minor centrifuge, England) for 10 minutes. Sera were used for assessment of kidney function. Kidneys from all the rats were cleared of any adherent tissues, weighed and fixed 10% formalin.

Determination of kidney function parameters

Assessment of kidney function was conducted using commercially available kit as described by the manufacturer. Parameters evaluated included bicarbonate, chloride, cholesterol, creatinine, potassium, sodium and urea.

Histological preparation

The tissues were dehydrated in ascending grades of alcohol (50% - 100%), impregnated and embedded in paraffin, sectioned at 5 μ m, mounted on glass slide, cleared in xylene, rehydrated in descending grades of alcohol (100% - 50%), and finally stained using hematoxylin and eosin. Tissue sections were viewed using Olympus biological microscope (CX23, Japan). All photomicrographs were produced at magnification of one hundred (×100).

Data analysis

Data were analyzed using GraphPad Prism version 9.2.0. Results from this research were presented as mean±SEM. *P*-values less than 0.05 were considered statistically significant.

RESULTS

Acute oral toxicity of the aqueous root extract of *S. guineense*

Administration of various doses of the extract in phase I of the study did not cause death or any observable signs of toxicity in the treated groups. Likewise, administration of the various doses in phase II did not result in mortality. However, at 5000 mgkg⁻¹ the rat stayed motionless and thereafter remained sluggish for about an hour.

Effects of the extract on body and kidney weights

Table 1 showed the body weight, body weight differences and kidney (absolute and relative) weight. Absolute kidney weight was obtained by weighing the kidneys directly using (AIPI-SS2) digital scale, GSS Scale (Suzhou) Co., Ltd. China. Relative kidney weight was obtained by dividing the absolute kidney weight with the body multiplied by 100. The body weights in all the groups showed no significant difference. Absolute kidney weights showed no significant difference among all the treated groups. However, relative kidney weights showed significant decrease. The relative kidney weight in the group administered with 10 mg/kg of the extract showed significant decrease (p<0.05) when compared with the control group. Likewise, the groups administered with 100 mg/kg of the extract also showed significant decrease (p<0.01) in relative kidney weight when compared with the control group.

Effects of the *S. guineense* aqueous root extract on renal functions

Results of renal functions were presented in Table 2. Cholesterol, creatinine, bicarbonate, chloride and potassium showed no significant changes in all the treated groups. However, urea in the group administered with 1000 mgkg⁻¹ increased significantly (p<0.05) when compared with the control group. Likewise, plasma sodium concentration also showed significant (p<0.05) increase in the groups administered with 10 mgkg⁻¹, 100 mgkg⁻¹ and 1000 mgkg⁻¹ when compared with the control group.

Effects of the *S. guineense* aqueous root extract on the histology of the kidney

Sections of kidneys from the control group and the groups administered with 10, 100 and 1000 mgkg⁻¹, as well as those administered with 1600, 2900 and 5000 mgkg⁻¹ were shown in Figure 1. Sections from the control group and the group administered with 10 mgkg⁻¹ showed intact normal renal corpuscle (RC), Bowman's capsule (BC) and renal tubule ($\stackrel{(l)}{\approx}$). Lymphocytes are indicated by letter (L). At doses of 1600, 2900 and 5000 mgkg⁻¹ renal tubules did not differ remarkably from the control group.

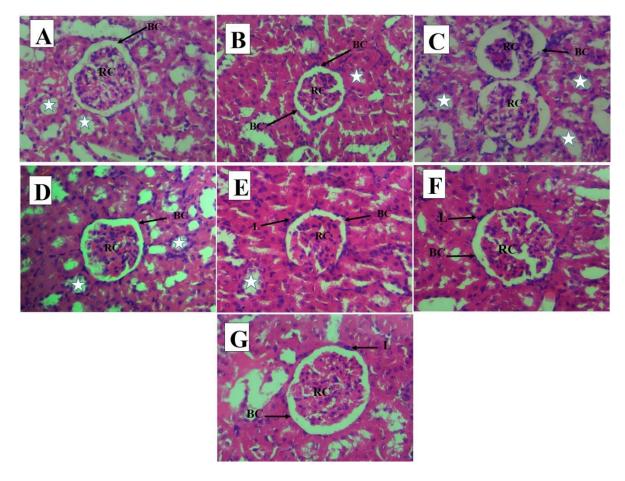


Figure 1: Sections from kidneys, plate (A) control received 0 mgkg⁻¹, plate (B) received 10 mgkg⁻¹, plate (C) received 100 mgkg⁻¹, plate (D) received 1000 mgkg⁻¹, plate (E) received 1600 mgkg⁻¹, plate (F) received 2900 mgkg⁻¹ and plate (G) received 5000 mgkg⁻¹. Sections from the treated groups showed no remarkable differences from the control group, except for the lymphocytes observed at doses of 1600 mgkg⁻¹, 2900 mgkg⁻¹ and 5000 mgkg⁻¹ (Haematoxylin and Eosin).

Table 1:Body and Kidney Weights

	Body Weight (g)			Kidney Weight (g)		
Doses (mgkg ⁻¹)	FBW	IBW	BWD	Relative (%)	Absolute (g)	
0	198.90±8.14	150.30±0.52	44.87±1.30	0.47±0.04	0.67±0.02	
10	192.27±15.44	147.40 ± 14.15	53.60±4.33	0.36±0.02*	0.68±0.02	
100	214.13±1.16	160.80±5.72	40.00±2.31	0.30±0.02**	0.65±0.04	
1000	202.67±3.93	162.57±1.70	46.60±8.66	0.31±0.02**	0.62±0.02	

IBM= Initial Body Weight; FBW=Final Body Weight; BWD= Body Weight Difference. *p<0.05; **p<0.01 compared with control

Table 2:Biomarkers of Renal Functions

	(mg/dL)						(mmol/L)	
Doses (mgkg ⁻¹)	Cholesterol	Creatinine	Urea	Bicarbonate	Chloride	Potassium	Sodium	
0	55.40±8.60	2.30±1.35	31.47±1.65	84.50±3.18	21.00±2.31	9.67±0.78	88.79±12.50	
10	42.70±7.33	0.71 ± 0.01	34.87±11.06	$103.50{\pm}10.10$	22.00±3.46	17.90 ± 11.74	116.30±3.06*	
100	60.47±5.51	0.60±0.06	42.87±4.34	82.00±1.73	19.00±1.16	6.75±0.49	111.70±8.20*	
1000	70.70±5.77	0.97±0.09	62.87±11.45*	88.50±4.91	28.50±3.75	6.07±0.32	114.17±0.26*	

*p<0.05 compared with control group

DISCUSSION

In this study, the absence of deaths at 5000 mg/kg of the extract is an indication of safety. Acute oral toxicity study of the aqueous root extract of *S. guineense* revealed that it is not lethal. This finding is in line with the work of Loha¹⁷. According to the United Nations¹⁸, the root extract of *S. guineense* has relatively low toxicity and belongs to category 5. Plant leaves extracts of *Pentaclethra macrophylla* and *Psidium guajava*, and that of *Spondias purpurea* also have oral LD₅₀ greater than 5000 mg/kg¹⁹. Substances or mixtures having oral median lethal dose greater than 5000 mg/kg are considered practically non-toxic or safe^{19, 20}.

Significant decrease in absolute kidney weight in all the treated groups suggests that the root extract of *S. guineense* might be toxic to the kidneys to some level. Sellers et al reported that decrease in organ weight is treatment related. Such a decrease of organ weight in toxicity study is one of the most important indicators of toxicity ^{21, 22}. Significant decrease in relative kidney weight in this study can be supported by the work of Abebe *et al.* where they reported decrease in relative organ weight of uterus and ovaries when methanolic leaves extract of *S. guineense* was administered to female rats ²³.

Decreased water excretion will go along with decreased sodium excretion; this implies increased reabsorption of water secondary to increased reabsorption of sodium in the renal tubules. This is suggestive of probable anti-diuretic effect of the plant. Such an increase of plasma sodium could cause high blood pressure and cardiovascular diseases ²⁴. Conditions such as acute renal injury, advanced chronic renal disease and glomerular diseases cause the kidneys to fail to perform their normal functions, and primarily cause high levels of sodium in the blood ²⁵.

Approximately 50% of the urea excreted by the kidneys in glomerular filtration is reabsorbed²⁶. Significant increase in plasma urea might be due to toxic effects on the renal tubules, renal parenchyma, and blockages in the urine outflow tract such as calculi, crystalluria, or other obstructions, in addition to glomerular alterations²⁷. Significant increase in the plasma concentration of urea in this research might be due to the toxic effects of the extract on the renal tubules or renal parenchyma. This finding is in line with the findings of Abebe²⁸. Ethanolic extract of *Chromolaena odorata*²⁹ and methanolic root extract of *Taraxacum officinale*³⁰ were also reported to have caused significant increase of

plasma urea. However, significant increase in plasma urea level might not be an indication of renal impairment because plasma creatinine levels in the treatment groups did not show any significant increase³⁰.

Histologically, no conspicuous differences were observed between the kidneys of the control and the treated groups. This finding is in line with the findings of Loha¹⁷ and Abebe *et al* ²⁸ who reported the absence of histologically remarkable differences between the kidneys in the control group and the treated groups.

CONCLUSION

The study revealed that oral administration of aqueous root extract of *S. guineense* caused decrease in relative kidney weight and increase in the plasma concentrations of sodium and urea in albino Wister rats. Therefore, it is recommended that the root of *S. guineense* should be consumed with caution.

Acknowledgements

We acknowledge the efforts of Mr. Ephraim Ayuba and Mr. Sunday Joseph Manye of Histology Laboratory, Department of Human Anatomy, Faculty of Basic Medical Sciences University of Maiduguri, for the assistance rendered during the sacrifice of the rats.

REFERENCES

- 1. Jones LR, Wright SJ, Gant TW. A critical review of microplastics toxicity and potential adverse outcome pathway in human gastrointestinal tract following oral exposure. Toxicology Letters. 2023;385: 51-60.
- Badou R, Yedomonhan H, Ewedje EB, Dassou G, Adomou A, Tossou M, *et al.* Floral morphology and pollination system of *Syzygium guineense* (Willd) DC. subsp. macrocarpum (Engl.) F. White (Myrtaceae), a subspecies with high nectar production. South African Journal of Botany. 2020; 131:462-7.
- 4. Negash L. A Selection of African Native Trees: Biology, Uses, Propagation and Restoration Techniques, Addis Ababa, Ethiopia. Legesse Negash, 2021.

- 5. Koval A, Pieme CA, Queiroz EF, Ragusa S, Ahmed K, Blagodatski A, *et al.* Tannins from Syzygium guineense suppress Wnt signaling and proliferation of Wnt-dependent tumors through a direct effect on secreted Wnts. Cancer Letters. 2018;435: 110-20.
- Uddin ABMN, Hossain F, Reza ASMA, Nasrin MS, Alam AHMK. Traditional uses, pharmacological activities, and phytochemical constituents of the genus *Syzygium*: A review. Food Science & Nutrition. 2022;10(6):1789-819.
- 7. Tadesse SA, Wubneh ZB. Antimalarial activity of Syzygium guineense during early and established Plasmodium infection in rodent models. BMC Complement Altern Med. 2017;17(1)
- 8. Ezenyi IC, Igoli JO. Antidiarrhoeal properties of *Syzygium guineense* leaf extract and identification of chemical constituents in its active column fractions. Journal of Complementary and Integrative Medicine. 2019 Jun 26;16(2)
- Ezuruike UF, Chieli E, Prieto JM. In Vitro Modulation of Glibenclamide Transport by Pglycoprotein Inhibitory Antidiabetic African Plant Extracts. Planta Med. 2019;85(11/12):987-96.
- 10. Oladosu IA, Aiyelaagbe OO, Afieroho OE. A Novel Normethylfriedelane-Type Isoprenoid from Syzygium guineense Stem Bark. Chem Nat Compd. 2018;54(1):112-6.
- Seely JC, Hard GC, Blankenship B. Kidney, In: Boorman's Pathology of the Rat: Reference and Atlas. Suttie AW. (ed.) 2nd edn. Academic Press, London, UK. 2018
- 12. Al-Kuraishy H, Al-Naimi M, Rasheed H, Hussien N, Al-Gareeb A. Nephrotoxicity: Role and significance of renal biomarkers in the early detection of acute renal injury. J Adv Pharm Technol Res. 2019;10(3):95.
- Inker LA, Titan S. Measurement and Estimation of GFR for Use in Clinical Practice: Core Curriculum 2021. American Journal of Kidney Diseases. 2021;78(5):736-49.
- Loffing J, Verrey F, Wagner CA. The kidneys matter. Pflugers Arch - Eur J Physiol. 2022;474(8):755-7.
- Dudas B. Human Histology: A Text and Atlas for Physicians and Scientists. Academic Press. London, UK. 2023
- Lorke D. A new approach to practical acute toxicity testing. Arch Toxicol. 1983 54(4):275-87.
- 17. Loha M, Mulu A, Abay SM, Ergete W, Geleta B. Acute and Subacute Toxicity of Methanol Extract of *Syzygium guineense*Leaves on the

Histology of the Liver and Kidney and Biochemical Compositions of Blood in Rats. Evidence-Based Complementary and Alternative Medicine. 2019; 2019:1-15.

- United Nations. Globally Harmonized System of Classification and Labelling of Chemicals: Acute Toxicity. 10th rev. edition, United Nations Publications, New York, USA 2023.
- Yamssi C, Payne VK, Noumedem Anangmo CN, Tateng Ngouateu A, Megwi L and Kuiate JR. Acute Toxicity of Pentaclethra macrophylla and Psidium guajava Use as Antiprotozoan Medicinal Plants. J Drug Discov Develop and Deliv. 2020; 6(1): 1037
- Oluwaiye J, Shehu A, Aliyu IM, Kwanashie HO, Anafi SB, Ibrahim A. Safety assessment of Spondias purpurea aqueous leaf extract (anacardiaceae): Acute and sub-chronic toxicity studies in Wistar rats. Journal of Current Biomedical Research 2023;3(1):741-67.
- Sellers RS, Mortan D, Michael B, Roome N, Johnson JK, Yano BL, *et al.* Society of Toxicologic Pathology Position Paper: Organ Weight Recommendations for Toxicology Studies. Toxicol Pathol. 2007;35(5):751-5.
- 22. Piao Y, Liu Y, Xie X. Change Trends of Organ Weight Background Data in Sprague Dawley Rats at Different Ages. J Toxicol Pathol. 2013;26(1):29-34.
- 23. Abebe MS, Asres K, Bekuretsion Y, Woldekidan S, Debebe E, Abebe A, *et al.* Toxic effect of Syzygium guineense ethanolic extract on female reproduction in rats: An evidence from a 10 week repeated-dose toxicity study. Heliyon. 2023;9(6): e17335.
- 24. Adamczak M, Wiecek A. Food Products That May Cause an Increase in Blood Pressure. Curr Hypertens Rep. 2020;22 (1). https://doi.org/10.1007/s11906-019-1007-y
- Ellison DH. Disorders of Extracellular Volume, In: Johnson RJ, Floege J, Tonelli M. (Eds.). Comprehensive Clinical Nephrology, 7th edn Lippincott Williams and Wilkins Pennsylvania, USA. 2019.
- 26. Nong Y, Wei X, Qiu H, Yang H, Yang J, Lu J, *et al.* Analysis of risk factors for severe acute kidney injury in patients with acute myocardial infarction: A retrospective study. Front Nephrol. 2023; 3:1047249.
- 27. Evans GO. Animal clinical chemistry: a practical handbook for toxicologists and biomedical researchers (2nd ed.). CRC Press/Taylor & Francis. New York, USA. 2009.
- 28. Abebe MS, Asres K, Bekuretsion Y, Abebe A, Bikila D, Seyoum G. Sub-chronic toxicity of ethanol leaf extract of Syzygium guineense on

the biochemical parameters and histopathology of liver and kidney in the rats. Toxicology Reports. 2021; 8:822-8.

- 29. Anyanwu S, Inyang IJ, Asemota EA, Obioma OO, Okpokam DC, Agu VO. Effect of ethanolic extract of Chromolaena odorata on the kidneys and intestines of healthy albino rats. Integrative Medicine Research. 2017;6(3):292-9.
- Bekhaled I, Benalia A, Mehida H, Meziani S, Tarfaoui L, Djjebar AA, *et al.* Evaluation of the Acute Toxicity of Dandelion (Taraxacum officinale) Roots. J Drug Delivery Ther. 2020;10(3):159-63.